

# What lives in and on the shell of Pāua? Epibionts on Haliotis iris from Kaikōura. http://www.ocean-wolf.com

Cost of epibiont to industry

Fouling organisms can be problematic for aquacultural infrastructure by adding weight to equipment, blocking pumps and increasing energy costs. Those organisms and shellinfesting species, like worms, sponges, and others, adversely affect the cultured species and reduce their market value. From the USA, we know that the mitigation of fouling organisms to shellfish aquaculture averages around 15% (21) million USD) of the annual cost. Depending on shellfish species, the cost of managing epibionts can increase to 30%

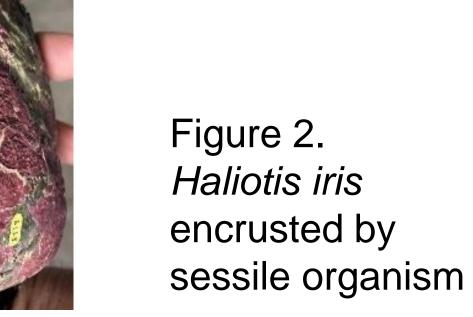
# Effects of epibionts on shelfish

R. Paul Wolf

Shell fouling and shell-infesting species (epibionts) can be detrimental to the shellfish's survival, growth and reproduction. With rising temperatures, effects like shell weakening (fig. 1), limited mobility, and increased weight caused by native and non-indigenous epibionts (fig. 2) will substantially increase energetic requirements for the shellfish while reducing survival and reproductive output <sup>8–13</sup>. Particular epibionts from foreign shores present further substantial threats to local shellfish populations as they can be vectors for other diseases and

#### of the final product 1-7.

Figure 1. Haliotis *iris* with an eroded shell





#### parasites <sup>14–17</sup>. Many knowledge gaps remain, not least because bio-fouler/mollusc interactions have not yet been studied in New Zealand. Gaps range from basic biology to interactions between epibionts and their combined impact on shellfish. Particular for abalone such as Haliotis iris (Gmelin, 1791) are studies limited to a few worm and sponge species and their distribution along our coastline. Given the commercial and cultural importance of paua, the lack of knowledge in epibiont and shellfish interaction is astonishing.

#### The work

Prior to being a taxonomist at Cawthron, I was involved in recovery projects in Kaikoura. During this time, I adopted methods in abalone shell cleaning from literature <sup>2,4</sup>. Colleagues and I carefully cleaned the shells off of 120 individuals that we brought in from the wild. The cleaning was done by gently scraping off encrusting species and covering holes in the shell with surf wax to kill boring worms. This procedure had no noticeable effects on the behaviour and general health of the abalone. Remarkably, the removal of epibionts reduced the drip weight of the individual by up to 30%, suggesting a significant load of epibiota was present. A few months after the cleaning, the individuals seemed to be in good physical condition.

# First results

A subsample of removed organisms was analysed for species composition and abundance. Eleven taxa were identified, with limpets and chitons being the commonest. Various sessile and mobile polychaete worms were abundant (Table 1), including several individuals of Polydora spp. (shell-infesting worms, fig. 4). With these first results, I hope to encourage further research into epibionts, parasite-host association and which affect these organisms have on shellfish. Such work provides a baseline for hatchery managers to know what sort of marine species they may be bringing into their hatcheries and will inform strategies to optimise broodstock health. As well as understand the association between epibiont species to the host organisms.

Figure 3. Haliotis iris after shell cleaning. Wholes caused by worms closed up with wax to suffocate the infesting organism



### REFERENCES

1. Adams, C. M., Shumway, S. E., Whitlatch, R. B. & Getchis, T. Biofouling in Marine Molluscan Shellfish Aquaculture: A Survey Assessing the Business and Economic Implications of Mitigation. J. World Aquac. Soc. 42, 242–252 (2011); 2. Watson, D. I., Shumway, S. E. & Whitlatch, R. B. Biofouling and the shellfish industry. Shellfish Saf. Qual. 317–337 (2009); 3. Fitridge, I., Dempster, T., Guenther,

J. & de Nys, R. The impact and control of biofouling in marine aquaculture: A review.

Biofouling 28, 649–669 (2012); 4. Heasman, M. & Savva, N. Manual for intensive hatchery production of abalone: Theory and practice for year-round, high density seed production of blacklip abalone (Haliotis rubra). (2007); 5. El-Menif, N. T., Guezzi, Y., Lahbib, Y., Ramdani, M. & Flower, R. Effects of Biogenic Concretions, Epibionts, and Endobionts on the Relative Growth of the Clam Venus verrucosa in Bizerta Lagoon, Tunisia. J. Shellfish Res. 27, 1–6 (2008); 6. Moreno, R. A., Neill, P. E. & Rozbaczylo, N. Native and non-indigenous boring polychaetes in Chile: A threat to native and commercial mollusc species. Rev. Chil. Hist. Nat. 79, 263–278 (2006); 7. Alfaro, A. C., Webb, S. C. & Barnaby, C. Variability of growth, health, and population turnover within mussel beds of Perna canaliculus in northern New Zealand. Mar. Biol. Res. 4, 376–383 (2008); 8. Webb, S. C. & Korrubel, J. L. Shell weakening in marine mytilids attributable to blue-green alga Mastigocoleus sp. (Nostochopsidaceae). J.SHELLFISH RES. 13, 11–17 (1994); 9. Cowen, R., Gawarkiewicz, G., Pineda, J., Thorrold, S. & Werner, F. Population Connectivity in Marine Systems: An Overview. Oceanography 20, 14–21 (2011); 10. Osawa, Y. & Tokeshi, M. Lunella shell damages and epibionts: what are inter-specific relations? Coast. Ecosyst. 5, 1–13 (2018); 11. Cummings, V. J., Smith, A. M., Marriott, P. M., Peebles, B. A. & Halliday, N. J. Effect of reduced pH on physiology and shell integrity of juvenile Haliotis iris (pāua) from New Zealand. PeerJ 2019, 1–23 (2019); 12. Rowley, A. F. et al. Marine ScienceThe potential impact of climate change on the infectious diseases of commercially important shellfish populations in the Irish Sea - a review. ICES J. of Marine Sci. (2014), 71, 741–759 (2014); 13. Hollowed, A. B., Planque, B. & Loeng, H. Potential movement of fish and shellfish stocks from the sub-Arctic to the Arctic Ocean. Fish. Oceanogr. 22, 355–370 (2013); 14. Dinamani, P. Potential disease-causing organisms associated with mantle cavity of Pacific oyster Crassostrea gigas in northern New Zealand. Dis. Aquat. Organ. 2, 55–63 (1986); 15. Huchette, S., Paillard, C., Clavier, J. & Day, R. Shell disease: Abnormal conchiolin deposit in the abalone Haliotis tuberculata. Dis. Aquat. Organ. 68, 267–271 (2006); 16. Brenner, M. et al. Bivalve aquaculture transfers in Atlantic Europe. Part B: Environmental impacts of transfer activities. Ocean Coast. Manag. 89, 139–146 (2014); 17. Costello, K. E., Lynch, S. A., O'Riordan, R. M., McAllen, R. & Culloty, S. C. The Importance of Marine Bivalves in Invasive Host–Parasite Introductions. Front. Mar. Sci. 8, 1–14 (2021).

Figure 4. Two individuals of *Polydora* spp. removed from pāua shells



Table 1. Subsample of taxa with individual count removed from *Haliotis iris* shells

| Species                          | count |
|----------------------------------|-------|
| Syllidae                         | 2     |
| Phyllodocidae                    | 5     |
| Dodecaceria berkeleyi Knox, 1972 | 1     |
| Perinereis spp.                  | 2     |
| Polydora spp.                    | 5     |
| Spirobis                         | 6     |
| Sabellida                        | 1     |
| Patelloida spp.                  | 57    |
| Chiton glaucus Gray, 1823        | 10    |
| Limpet                           | 1     |

Acknowledgments: I am great full for the help of Shawn Gerrity & Jason Ruawai, and the support by Ministry of Primary Industries— earthquake relief fund; Paua Industry Council and Paua Mac3